



Review Article

Critical review on analytical detection of first line and second line anti tubercular agent by various modern analytical method

TS Basuri^{1*}, Ranjit Mohapatra²

¹Mayurbhanj Medical Academy, Baripada, Odisha, India

²Utkal University, Bhubaneswar, Odisha, India



ARTICLE INFO

Article history:

Received 14-03-2024

Accepted 21-05-2024

Available online 12-06-2024

Keywords:

Tuberculosis

First line agents

Second line agents

Analytical detection

HPLC

ABSTRACT

Tuberculosis is a chronic inflammatory, granulomatous bacterial infectious disease caused by *Mycobacterium tuberculosis*. About 10 million people worldwide are ill with MTB in 2018, 5.7 million are males, 3.2 million are females and 1.1 million are infants, 1.6 million died from the disease. This article includes epidemiology, classification, pathogenesis, diagnosis and treatment of Tuberculosis. It includes the drug profile of antitubercular agents such as isoniazid, pyrazinamide, ethambutol, rifampicin, paraminosalicylic acid, thiacetazone, ethionamide, kanamycin, amikacin, cycloserine, viomycin, morphazinamide and some newer drugs such as ofloxacin, ciprofloxacin, clarithromycin and rifabutin. It contains analytical detection methods of antitubercular agents by HPLC.

This is an Open Access (OA) journal, and articles are distributed under the terms of the [Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License](https://creativecommons.org/licenses/by-nc-sa/4.0/), which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: reprint@ipinnovative.com

1. Introduction

Tuberculosis (TB): Tuberculosis is a chronic granulomatous inflammatory infectious bacterial disease caused by *Mycobacterium tuberculosis*, which is most commonly affect the lungs.¹

1.1. Epidemiology of TB

MTB has very ancient origins: it has lasted more than 70,000 years and actually infects almost 2 billion people worldwide in 2016, with about 10.4 million new cases of TB annually, about 33.33% of the world's population carries TB bacillus and is at risk of developing active disease.²

Approximately 10 million people worldwide are infected with MTB in 2018, 5.7 million men, 3.2 million women and 1.1 million girls, 1.6 million people expired from the disease currently available for treating TB, it remains incompetent, taking 6 to 9 months to cure the drug-susceptible variant and

up to 2.5 years to cure MDR-TB.³

1.2. Classification of Tuberculosis (TB 4):

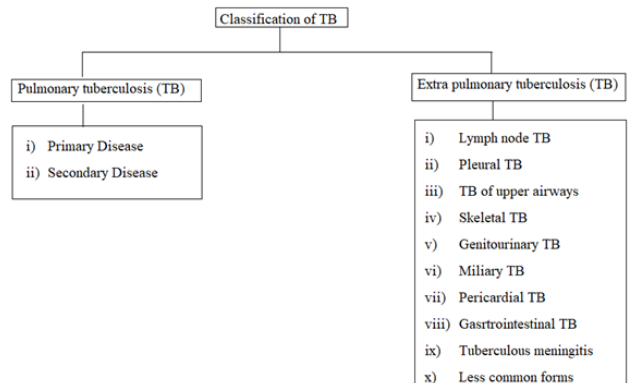


Figure 1: Classification of tuberculosis

* Corresponding author.

E-mail address: tbasuri@gmail.com (T. S. Basuri).

1.3. TB Pathogenesis

1. TB is an infectious disease caused by *M. Tuberculosis* usually affects the lungs.⁴
2. *M. Tuberculosis* is transmitted to atmosphere as airborne droplets, owing to runny noses of individuals with pulmonary tuberculosis (TB). Transmission done due to breathing of nuclei of these droplets which goes from the nasal cavities to upper respiratory tract, bronchi and then lastly goes to the alveoli of lungs.⁵
3. First on *M. Tuberculosis* enters the alveoli and is swallowed by alveolar macrophages that cause a large ratio of inhaled tuberculosis bacilli to be destroyed.⁶
4. The minor unchanged ratio reproduces inside the macrophages and is liberated upon death of the macrophages.⁷
5. Approximately within 2 to 8 weeks,⁸ an immune defensive mechanism is activated which permits leukocytes to extinguish major proportion of the tubercle bacilli. The encapsulation by the leukocytes leads to the barrier formation around the tubercle bacilli developing a granuloma.⁷
6. Once if it goes inside the hindrance shell, the tubercular bacilli are to be under control and it establish a stage of latent tuberculosis infection (LTBI). Persons will not give any indications of TB and infection are unable to spread.⁹
7. On other hand, if the defensive mechanism is unable to keep the tubercular bacilli under control, then bacilli are quickly reproduced which leads to a development from latent tuberculosis infection to tuberculosis TB.¹⁰

1.4. Diagnosis of TB^{11,12}

1. The medical history of patients
2. Physical test
3. *M. Tuberculosis* test
4. Chest X-ray
5. Bacteriologic examination

1.5. Treatment of tuberculosis:^{13,14}

Anti-tubercular agent: These are the medicines which are used for the tuberculosis treatment.

- Drug profile of antitubercular agents Table 1
- Analytical Detection of Antitubercular Agents
- First line antitubercular drugs Table 1
- Second line antitubercular drugs: Table 2
- New antitubercular drugs Table 4

2. Conclusion

Tuberculosis is a prolonged bacterial infection caused by *Mycobacterium tuberculosis*, characterised by the development of granulomas in infected tissues and by hypersensitivity mediated by the cells. About 10

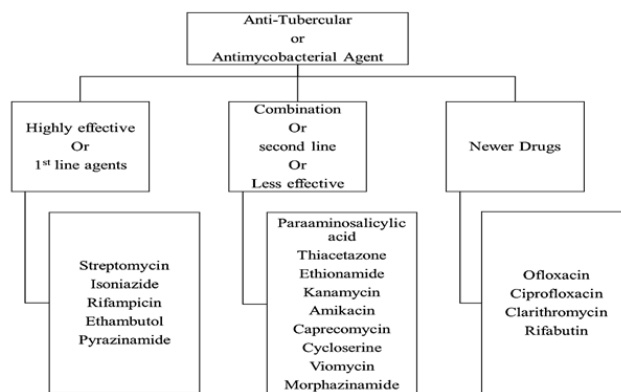


Figure 2: Class of TB drugs

Table 1	First line ant tubercular agents	Strature	Second line antitubercular agent	New drugs used in tuberculosis
1		1		1
2		2		2
3		3		3
4		4		4
		5		5
		6		
		7		

Diagram 1: Table 1+ Structure

million people worldwide became contaminated with MTB in 2018, 5.7 million men, 3.2 million women and 1.1 million kids, 1.6 million die from illness. This article includes epidemiology, classification, pathogenesis, diagnosis and treatment of Tuberculosis. It includes the drug profile of antitubercular agents such as isoniazid, pyrazinamide, ethambutol, rifampicin, paraminosalicylic acid, thiacetazone, ethionnamide, kanamycin, amikacin, cycloserine, viomycin, morphazinamide and fewer new mwdicines include ofloxacin, ciprofloxacin, clarithromycin and rifabutin. It contains analytical detection method of

Table 1: Drug description of antitubercular agent

Sr. no.	Drug	Mechanism of action
1.		
i)	Isoniazide ^{15,16}	Isoniazid interferes with the cell wall creation by inhibiting the synthesis of mycolic acid.
ii)	Pyrazinamide ^{17,18}	Pyrazinamide is a synthetic pyrazinoic acid amide derivative having bactericidal property. Pyrazinamide is mainly dynamic against gradually multiplying intracellular bacilli by an unfamiliar mechanism of action. Its bactericidal action is based on the bacterial pyrazinamidase, which eliminates the amide group to generate active pyrazinoic acid.
iii)	Ethambutol ^{19–21}	Ethambutol prevent the transfer of mycolic acid in M. tuberculosis cell wall. Which leads to the weakening of cell wall resulting into cell death.
iv)	Rifampicin ^{22,23}	Rifampicin works by blocking DNA dependent RNA polymerase, which results in depletion of synthesis of RNA and death of cell.
2.		
i)	Paraminosalicylic acid ²⁴	Aminosalicylic acid works by two route. 1 st - It inhibit folic acid production. Aminosalicylic acid bind to pteridine synthetase enzyme which is use for synthesis of folic acid. As bacteria is not able to use exterior source of folic acid leads to slow down the multiplication and growth of cell. 2 nd - It may hinder the production of the cell wall constituent, mycobactin, which leads to dropping iron uptake by M. tuberculosis.
ii)	Thiacetazone ²⁵	It is a bacteriostatic agent. Exact mechanism of thiacetazone is unknown. It can inhibit the synthesis of mycolic acid in mycobacterium tuberculosis leading to cell wall weakening which results in cell death.
iii)	Ethionamide ²⁶	Ethionamide is a derivative of nicotinamide. Actual mechanism of ethionamide is mysterious. It might prevent the creation of mycolic acid which leads to bacterial cell wall interruption and cell lysis.
iv)	Kanamycin ²⁷	Kanamycin is an antibiotic class of the aminoglycoside. Aminoglycosides function by binding to the bacterial ribosomal subunit 30S, producing misinterpretation of t-RNA, Leads to bacteria is not able to synthesize protein which are essential for growth of cell.
v)	Amikacin ²⁸	Amikacin fixes to bacterial 30S ribosomal subunits and affects with mRNA binding lead to inhibit protein synthesis which are necessary for its growth.
vi)	Cycloserine ²⁹	Cycloserine blocks peptidoglycan formation, allows the cell wall to collapse and contributes to cell death.
vii)	Viomycin ^{30,31}	Viomycin restricts translocation which obstructs the synthesis of proteins, leading to death of bacterial cells.
3.		
i)	Ofloxacin ³²	Ofloxacin is a fluoroquinolone antibiotic. Ofloxacin inhibits bacterial topoisomerase II and topoisomerase IV mechanism, which are participating in duplication and repair of DNA, leads to cell death.
ii)	Ciprofloxacin ³³	Ciprofloxacin is a synthetic broadspectrum fluoroquinolone antibiotic. Ciprofloxacin inhibits bacterial DNA gyrase mechanism, leads to obstruction of DNA replication, resulting in cell death.
iii)	Clarithromycin ³⁴	Clarithromycin attach to 50S subunit of ribosome and hunder protein production in bacteria which is important for its growth.
iv)	Rifabutin ³⁵	Rifabutin hinders bacterial DNA-dependent RNA polymerase, which inhibit transcription or RNA synthesis. Which leads to inhibition of protein synthesis which are essential for cell growth.

Table 2: Analytical detection of first line antitubercular agents by HPLC

Sr. no	Method	Mobile phase	Stationary phase	Flow rate (ml/min)	Wavelength (nm)	Retention time (min)
i)	Isoniazide ³⁶	Ethanol: water: 1%acetic acid (5.3:93.7:1, v/v/v)	C-18 column	1.5	265	2.28
	Isoniazide ³⁷	Water: methanol (85:15, v/v)	C-8 column (250 x 4.6mm,5m)	1.2	274	4.1
ii)	Pyrazinamide ³⁸	0.02M Potassium dihydrogen phosphate (pH 2.6): acetonitrile (98:2, v/v)	C-18 column (25 cm × 4.6 mm, 5 μm)	1	268	8.4
iii)	Ethambutol ³⁹	Methanol: water: glacial acetic acid (70:30:0.2, v/v/v)	C-18 column	1	210	29.26
iv)	Rifampicin ⁴⁰	Methanol: acetonitrile: (0.075 M) monopotassium phosphate: (1.0 M) citric acid (28:30:38:4, v/v)	C-18 column (150 × 4.6 mm, 5m)	2	254	2.91

Table 3: Analytical detection of second line antitubercular agents by HPLC

i)	Paraminosalicylic acid ⁴¹	20 mM phosphate buffer, 20mM tetrabutylammonium hydrogen sulphate and 16% (v/v) methanol adjusted to pH 6.8	C-18 column (50 mm × 4.6 mm, 5 μm)	1	233	2.783
ii)	Thiacetazone ⁴²	Water: acetonitrile (92.5:7.5, v/v)	Internal surface reversed-phase (ISRP) mixed-functional phenyl column (Capcell Pak, 50x4.6 mm, 5 μm)	1	322	10.9
iii)	Ethionamide ⁴³	Acetonitrile: phosphate buffer (75:25, v/v)	C-18 column (250×4.6mm, 5μm)	1.5	291	3.8
	Ethionamide ⁴⁴	Methanol: water (40:60, v/v)	C-18 column (250 × 4.6 mm, 5 μm)	1	275	5.467
iv)	Kanamycin ⁴⁵	22 mM disodium 1,2-ethanedisulfonate and 5 mM sodium octanesulfonate in a water-acetonitrile mixture (80:20, v/v)	C-18 column (40 × 8 mm,10 μm)	1.5	Ex: 351 Em: 440	9
	Kanamycin ⁴⁶	Acetonitrile:0.1 M sodium acetate buffer (pH 5.0; 25:75, v/v)	C-18 column (250 x 4.6 mm, 5 μm)	2	330	19.5
v)	Amikacin ⁴⁷	Acetonitrile:0.1 M sodium acetate buffer (pH 5.0; 25:75, v/v)	C-18 column (250x4.6 mm, 5 μm)	2	330	8.7
	Amikacin ⁴⁸	Methanol:acetonitrile: acetate buffer (75:20:05 v/v)	C-18 column (250 mm x 4.6 mm, 5 μm)	1	212	4.61
vi)	Cycloserine ⁴⁹	20mM Sodium hydrogen phosphate: acetonitrile (95:05, v/v)	Agilent Zorbax SB phenyl column (250 × 4.6 mm, 5 μm)	1	335	L- 10.4 D- 11.8
	Cycloserine ⁵⁰	10 mM Phosphate buffer: acetonitrile (90:10, v/v)	Atlantis T3, (150 × 4.6 mm, 3 μm)	0.4	240	5.1

Table 4: Analytical detection of new antitubercular agents by HPLC

i)	Ofloxacin ⁵¹	Acetonitrile: Buffer (35:65, v/v)	C-8 column (250 cm × 4.6 mm, 5 μm)	1.5	315	10
	Ofloxacin ⁵²	Triethylamine: acetonitrile :0.3% o-phosphoric acid ((0.02:20:80, v/v/v)	C-18 column (250 × 4.6 mm, 5 μm)	1	290	6.15
	Ofloxacin ⁵³	0.03M Potassium dihydrogen phosphate: methanol (30:70, v/v)	C-18 column (150 × 4 mm, 4 μm)	1	294	5.17
ii)	Ciprofloxacin ⁵⁴	Phospahte buffer (2.7 pH): Acetonitrile (77:23, v/v)	C-18 column (250 x 4.6 mm, 5 μm)	1.5	277	3.26
	Ciprofloxacin ⁵⁵	2% Acetic acid: acetonitrile (84:16, v/v)	C-18 column (150 × 4.6 mm, 5 μm)	1	280	6.5
	Ciprofloxacin ⁵⁶	5% Acetic acid: acetonitrile: methanol (90:5:5, v/v/v)	C-18 column (150 × 6 mm, 5 μm)	1	280	12
	Ciprofloxacin ⁵⁷	Water: acetonitrile: triethylamine (80:20:0.3, v/v/v)	C-18 column (125 x 4 mm, 5 μm)	1	279	2.4
C	Ciprofloxacin ⁵⁸	0.025M Orthophosphoric acid (3 pH): methanol (60:40, v/v)	C-18 column (125x4mm, 5μm)	2	278	1.75
	Ciprofloxacin ⁵⁹	0.1M Potassium dihydrogen phosphate: acetonitrile (80:20, v/v)	C-18 column (250 mm × 4.6 mm, 10)	1.5	276	5.15
iii)	Clarithromycin ⁶⁰	Acetonitrile: methanol:0.04 M phosphate buffer (pH 6.9) (52:9:39, v/v)	Perkin-Elmer Spheri-5 cyano column (100 × 4.6 mm, 5 μm)	1	ECD	9.2
	Clarithromycin ⁶¹	0.05M Phosphate buffer (3.2 pH): acetonitrile (50:50, v/v)	C-18 column (250 × 4.6 mm, 5 mm)	1	205	2.21
	Clarithromycin ⁶²	Acetonitrile: methanol: potassium dihydrogen phosphate buffer (7.5 pH) (40:6:54, v/v)	C-8 column (125 × 4.0 mm)	1.5	Amperometric	4.8 detector
	Clarithromycin ⁶³	Acetonitrile: formic acid: water: trifluoroacetic acid (70:15:14.9:0.1, v/v)	C-18 column (250x4.6 mm, 5 μm)	1	ELSD	4.7
	Clarithromycin ⁶⁴	Acetonitrile: phosphate buffer (11 pH) (60:40, v/v)	Asahipak Shodex ODP-50 4E column (250 mm × 4.6 mm, 5 μm)	1	210	6.46
	Clarithromycin ⁶⁵	Acetonitrile: 0.035 M potassium dihydrogen phosphate (pH 4.4) (55: 45, v/v)	C-18 column (150× 4.6 mm, 5 μm)	0.6	210	4.1
iv)	Rifabutin ⁶⁶	50mM Phospahte buffer (4.2 pH): acetonitrile (53:47, v/v)	C-18 column (250 x 4.6mm, 5m)	1.2	265	8.5
	Rifabutin ⁶⁷	Methanol: Water (75:25, v/v)	C-8 column (250 × 4.6 mm, 5 μm)	1	240	5.5
	Rifabutin ⁶⁸	Acetonitrile + Methanol (1:1): Water (75:25, v/v)	C-18 column (250 x 4.6mm, 5m)	1	242	5.3

antitubercular agents by HPLC. From the study we can conclude that there are various analytical detection methods are available. The current work contains compilation of the analytical detection method antitubercular agents by HPLC.

3. Source of Funding

None.

4. Conflict of Interest

None.

References

- Shang S, Siddiqui S, Bian Y, Zhao J, Wang CR. Nonclassical MHC Ib-restricted CD8+ T cells recognize mycobacterium tuberculosis-derived protein antigens and contribute to protection against infection. *PLoS pathogens*. 2016;12(6):e1005688.
- Barberis I, Bragazzi NL, Galluzzo L, Martini M. 2017.
- World Health Organization. *World Health Organization World malaria report*. 2015;.
- Floyd K, Pantoja A. Financial resources required for tuberculosis control to achieve global targets set for 2015. *Bulletin of the World Health Organization*. 2008;86:568–76.
- Kay A, Barry PM, Annambhotla P, Greene C, Cilnis M, Chin-Hong P, et al. Solid Organ Transplant-Transmitted Tuberculosis Linked to a Community Outbreak-California. *MMWR Morbidity and mortality weekly report*. 2015;66:801–801.
- Schluger NW. 2005.
- Smith I. 2003.
- Haley CA. 2013.
- Raviglione M, Sulis G. 2016.
- Donald PR, Marais BJ, Barry CE. 2010.
- Golub JE, Bur S, Cronin WA, Gange S, Baruch N, Comstock GW, et al. 2006.
- Agyeman AA, Ofori-Asenso R. Tuberculosis-an overview. *Journal of Public Health and Emergency*. 2017;1(7).
- Radloff J, Heyckendorf J, Merwe LVD, Carballo S, Reiling P, Richter N, et al. Mycobacterium Growth Inhibition Assay of Human Alveolar Macrophages as a Correlate of Immune Protection Following Mycobacterium bovis Bacille Calmette-Guérin Vaccination. *Frontiers in immunology*. 2018;9:1708–1708.
- Maher D, Chaulet P, Spinaci S, Harries A. 1997.
- Timmins GS, Deretic V. 2006.
- Weber WW, Hein DW. *Clinical pharmacokinetics of isoniazid Clinical pharmacokinetics*. 1979;4(6):401–423.
- Zhang Y, Mitchison D. 2003.
- Shi W, Zhang X, Jiang X, Yuan H, Lee JS, Barry CE, et al. Pyrazinamide inhibits trans-translation in Mycobacterium tuberculosis. *Science*. 2011;333(6049):1630–1632.
- Takayama K, Kilburn JO. Inhibition of synthesis of arabinogalactan by ethambutol in Mycobacterium smegmatis. *Antimicrobial agents and chemotherapy*. 1989;33(9):1493–1502.
- Place VA, Thomas JP. 1963.
- Sreevatsan S, Stockbauer KE, Pan XI, Kreiswirth BN, Moghazeh SL, Jacobs WR, et al. Ethambutol resistance in Mycobacterium tuberculosis: critical role of embB mutations. *Antimicrobial agents and chemotherapy*. 1997;41(8):1677–81.
- Telenti A, Imboden P, Marchesi F, Matter L, Schopfer K, Bodmer T, et al. 1993.
- Acocella G. Clinical pharmacokinetics of rifampicin. *Clinical pharmacokinetics*. 1978;3(2):108–135.
- Ibrahim A, Boutros A, McDougall JB. Chemotherapy in a Cairo Chest Clinic: A preliminary report on the methods adopted in assessing the value of isoniazid, streptomycin and paraaminosalicylic acid in 122 cases of pulmonary tuberculosis. *British Journal of Tuberculosis and Diseases of the Chest*. 1955;49(1):38–49.
- Alahari A, Trivelli X, Guérardel Y, Dover LG, Besra GS, Sacchettini JC, et al. Thiacetazone, an antitubercular drug that inhibits cyclopropanation of cell wall mycolic acids in mycobacteria. *PLoS One*. 2007;2(12).
- Johansson K, King DS, Schultz PG. Studies on the mechanism of action of isoniazid and ethionamide in the chemotherapy of tuberculosis. *Journal of the American Chemical Society*. 1995;117(17):5009–5019.
- Misumi M, Tanaka N. 1980.
- Alangaden GJ, Kreiswirth BN, Aouad A, Khetarpal M, Igno FR, Moghazeh SL, et al. Mechanism of resistance to amikacin and kanamycin in Mycobacterium tuberculosis. *Antimicrobial agents and chemotherapy*. 1998;42(5):1295–1302.
- Prosser GA, Carvalho LPD. 2013.
- Stanley RE, Blaha G, Grodzicki RL, Strickler MD, Steitz TA. 2010.
- Holm M, Borg A, Ehrenberg M, Sanyal S. Molecular mechanism of viomycin inhibition of peptide elongation in bacteria. *Proceedings of the National Academy of Sciences*. 2016;113:978–83.
- Sun Z, Zhang J, Zhang X, Wang S, Zhang Y, Li C. Comparison of gyrA gene mutations between laboratory-selected ofloxacin-resistant Mycobacterium tuberculosis strains and clinical isolates. *International journal of antimicrobial agents*. 2008;31(2):115–136.
- Ciccione R, Mariani F, Cavone A, Persichini T, Venturini G, Ongini E, et al. Inhibitory effect of NO-releasing ciprofloxacin (NCX 976) on Mycobacterium tuberculosis survival. *Antimicrobial agents and chemotherapy*. 2003;1(7):2299–302.
- Bosne-David S, Barros V, Verde SC, Portugal C, David HL. Intrinsic resistance of Mycobacterium tuberculosis to clarithromycin is effectively reversed by subinhibitory concentrations of cell wall inhibitors. *Journal of Antimicrobial Chemotherapy*. 2000;46(3):391–396.
- Barluenga J, Aznar F, García AB, Cabal MP, Palacios JJ, Menéndez MA. 2006.
- Nguyen C.
- Hk AK, GR. Simple and rapid method for simultaneous determination of isoniazid and acetyl isoniazid in urine by HPLC. *Asian Journal of Biomedical and Pharmaceutical Sciences*. 2014;4(34):46–46.
- Conte JE, Lin E, Zurlinden E. High-performance liquid chromatographic determination of pyrazinamide in human plasma, bronchoalveolar lavage fluid, and alveolar cells. *Journal of chromatographic science*. 2000;38(1):33–40.
- Yan M, Guo T, Song H, Zhao Q, Sui Y. Determination of ethambutol hydrochloride in the combination tablets by precolumn derivatization. *Journal of chromatographic science*. 2007;45(5):269–272.
- Liu J, Sun J, Zhang W, Gao K, He Z. HPLC determination of rifampicin and related compounds in pharmaceuticals using monolithic column. *Journal of pharmaceutical and biomedical analysis*. 2008;46(2):405–414.
- Vasbinder E, Weken GVD, Heyden YV, Baeyens WR, Debunne A, Remon JP, et al. Quantitative determination of p-aminosalicylic acid and its degradation product m-aminophenol in pellets by ion-pair high-performance liquid chromatography applying the monolithic Chromolith Speedrod RP-18e column. *Biomedical Chromatography*. 2004;18(1):55–63.
- Song D, Wientjes MG, Au JL. Isocratic high-performance liquid chromatographic determination of thiacetazone by direct injection of plasma into an internal surface reversed-phase column. *Journal of Chromatography B: Biomedical Sciences and Applications*. 1997;690(1-2):289–94.
- Madni AU, Ahmad M, Akhtar N, Ashraf M, Shuja ZA. An improved HPLC method for the determination of ethionamide in serum. *Journal of the Chemical Society of Pakistan*. 2008;30(3):449–52.
- Rahade P, Sonawane S, Bhalerao A, Kshirsagar S. Development of a Validated RP-HPLC Method for Estimation of Ethionamide in Spiked Human Plasma with UV Detection. *Asian Journal of Research in Pharmaceutical Science*. 2016;6(4):230–234.
- Kubo H, Kobayashi Y, Nishikawa T. 1985.
- Korany MA, Haggag RS, Ragab MA, Elmallah OA. Liquid chromatographic determination of amikacin sulphate after pre-column

- derivatization. *Journal of chromatographic science*. 2014;52(8):837–884.
47. Teja GS, Gurupadayya BM, Sairam KV. Analytical method development and validation of amikacin in pure and marketed formulation using HPLC. *INTERNATIONAL JOURNAL OF PHARMACEUTICAL SCIENCES AND RESEARCH*. 2018;9(10):4382–4388.
 48. Korany MA, Haggag RS, Ragab MA, Elmallah OA. Liquid chromatographic determination of amikacin sulphate after pre-column derivatization. *Journal of chromatographic science*. 2014;52(8):837–884.
 49. Karthikeyan K, Arularasu GT, Ramadhas R, Pillai KC. Development and validation of indirect RP-HPLC method for enantiomeric purity determination of d-cycloserine drug substance. *Journal of pharmaceutical and biomedical analysis*. 2011;54(4):850–854.
 50. Kumar AH, Polisetty AK, Sudha V, Vijayakumar A, Ramachandran G. A selective and sensitive high performance liquid chromatography assay for the determination of cycloserine in human plasma. *Indian Journal of Tuberculosis*. 2018;65(2):118–141.
 51. Patel SK, Smith AA, Amuthalakshmi S, Gandhi VN, Manavalan R. Analytical method development and validation of Ofloxacin eye drop by HPLC. *J Curr Chem Pharm Sc*. 2011;1(1):59–64.
 52. Maslarska V, Tsvetkova B, Peikova L, Bozhanov S. RP-HPLC Method for Simultaneous Determination of Metronidazole and Ofloxacin in Synthetic Mixture. *InCUBU International Conference Proceedings*. 2016;4:900–905.
 53. Amini M, Abdi K, Darabi M, Shafiee A. Determination of ofloxacin in plasma by HPLC with UV detection. *Applied Sci*. 2005;5(9):1655–1655.
 54. Vella J, Busuttill F, Bartolo NS, Sammut C, Ferrito V, Serracino-Inglott A, et al. A simple HPLC-UV method for the determination of ciprofloxacin in human plasma. *Journal of Chromatography B*. 2015;989:80–85.
 55. Wu SS, Chein CY, Wen YH. Analysis of ciprofloxacin by a simple high-performance liquid chromatography method. *Journal of chromatographic science*. 2008;46(6):490–495.
 56. Kamberi M, Tsutsumi K, Kotegawa T, Nakamura K, Nakano S. 1998.
 57. Kassab NM, Singh AK, Kedor-Hackmam ER, Santoro MI. 2005.
 58. Ali SA, Mmuo CC, Abdulraheem RO, Abdulkareem SS, Alemika ET, Sani MA, et al. High performance liquid chromatography (HPLC) Method development and validation indicating assay for ciprofloxacin hydrochloride. *Journal of Applied Pharmaceutical Science*. 2011;1(8):239–239.
 59. Emami J, Rezazadeh M. A simple and sensitive high-performance liquid chromatography method for determination of ciprofloxacin in bioavailability studies of conventional and gastroretentive prolonged-release formulations. *Advanced biomedical research*. 2016;5.
 60. Niopas I, Daftsios AC. Determination of clarithromycin in human plasma by HPLC with electrochemical detection: validation and application in pharmacokinetic study. *Biomedical Chromatography*. 2001;15(8):507–515.
 61. Gangishetty S, Verma S. 2013.
 62. Foroutan SM, Zarghi A, Shafaati A, Madadian B, Abolfathi F. Rapid high-performance liquid chromatographic method for determination of clarithromycin in human plasma using amperometric detection: Application in pharmacokinetic and bioequivalence studies. *Iranian journal of pharmaceutical research: IJPR*. 2013;12:65–65.
 63. Tzouganaki Z, Koupparis M.
 64. Habibi B, Ghorbel-Abid I, Lahsini R, Hassen B, Trabelsi-Ayadi DC, M. Development and validation of a rapid HPLC method for multiresidue determination of erythromycin, clarithromycin, and azithromycin in aquaculture fish muscles. *Acta Chromatographica*. 2019;31(2):109–121.
 65. Alam MM, Hossain MS, Bhadra S, Kumar U, Rouf AS. Development and validation of RP-HPLC method for quantitation of clarithromycin in matrix tablet dosage form. *Dhaka University Journal of Pharmaceutical Sciences*. 2017;30(1):69–75.
 66. Kumar AH, Sudha V, Ramachandran G. Simple and rapid liquid chromatography method for determination of rifabutin in plasma. *SAARC Journal of Tuberculosis*. 2012;9(2):26–35.
 67. Patil YD, Banerjee SK. 2012.
 68. Bartels H, Bartels R. Determination of rifabutin by high-performance liquid chromatography using on-line concentration and column switching. *Journal of Chromatography B: Biomedical Sciences and Applications*. 1996;686(2):235–275.

Author biography

TS Basuri, Associate Professor

Ranjit Mohapatra, Assistant Professor

Cite this article: Basuri TS, Mohapatra R. Critical review on analytical detection of first line and second line anti tubercular agent by various modern analytical method. *IP Int J Comprehensive Adv Pharmacol* 2024;9(2):91-97.